

Instructions for Use

CIMmultus™ SDVB 4000 mL cGMP Compliant Monolithic Column (2 µm channels)

CIM Convective Interaction Media®
934.9001-2



SARTORIUS

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1. About These Instructions for Use

These instructions are part of the device. They apply to the device product number indicated on the cover page.

1.1. Accompanying Documents

Column integrity test



2. Safety

⚠ WARNING

Denotes a hazard that may result in death or severe injury if it is not avoided.

⚠ CAUTION

Denotes a hazard that may result in moderate or minor injury if it is not avoided.

NOTICE

Denotes a hazard that may result in property damage if it is not avoided.

2.1. Intended Use

CIMmultus™ Monoliths are reusable chromatography devices for scalable high-resolution purification of complex biological samples. Inside the custom designed housing is a single-piece stationary phase with homogeneous channel size and surface chemistry. Without the need for column packing, CIMmultus™ Monoliths are ready for use after sanitisation and integrity test.

The backbone of SDVB column is highly hydrophobic with a strong affinity for hydrophobic or less polar compounds. It enables fast separation of hydrophobic analytes under reverse phase conditions. The following information is being provided to ensure proper product care and optimal product performance.

2.2. Safety Note

Follow the guidelines in this Instructions for Use. Improper use may result in malfunction, personal injury, or damage of the product or material. Follow safety instructions, wear gloves, safety glasses, and a lab coat during operation.

3. Technical Data

Column chemistry	SDVB (reverse phase; poly(styrene-co-divinylbenzene))
Channel radius	1050 nm (950 nm - 1150 nm)
Support matrix	poly(styrene-co-divinylbenzene)
Monolith dimensions	Outer diameter: 300 mm; inner diameter: 255 mm; length: 205 mm; bed volume (CV): 4000 mL
Connector	TC 1 in. (25 mm), 8 mm ID bore
Ligand density	N.D.
Operating flow rates	Up to 1 CV/min 4000 mL/min 135 cm/h. Do not go below 0.1 CV/min
Maximum pressure	0.7 MPa, 7 bar, 100 psi
Operating temperature	4 °C (39 °F) to 40 °C (104 °F)
Chemical stability	The PS-DVB monolith is highly stable across the entire pH range. It can tolerate common organic solvents (acetonitrile, methanol, ethanol). Exposure to aqueous mobile phases should be kept to a minimum. Avoid rapid changes from aqueous to organic mobile phase. A linear gradient or shallow steps should be used.
Recommended pH	Working range 2-13, cleaning in place 1-14
Storage conditions	2 °C (36 °F) to 25 °C (77 °F); 20 % ethanol
Shelf life	1 year

The linear flow rate can be calculated with the following equation and supporting data, which is available in the Technical Data.

$$\text{Average linear velocity, } u_{av} = \frac{F}{\pi \times L} \frac{\ln\left(\frac{D_o}{D_i}\right)}{(D_o - D_i)}$$

F is the flow rate in mL/min, Do and Di are the outer and inner diameter of the column and L is the column length.

4. Device Overview | Description

The housing of this CIMmultus™ column is made of epoxy thermoset material. Its surface is coated pinhole-free with biocompatible (USP Class VI) Parylene C.

NOTICE

Do not expose the column housing to pure acetone.

5. Installation

Remove the product from its shipping box or crate and place on a flat surface. Carefully inspect the product for any damage that may have occurred during shipping. Immediately report any such damage to your vendor and the courier. The product is shipped in the designated storage solution at ambient temperature and should be stored upon receiving as stated under Technical Data.

NOTICE

Larger columns are shipped in a wooden crate, and a suitable stand is provided in the packaging. The columns have either a stand (400 | 800 mL columns) or wheels (4 | 8 | 40 L columns). Place them in an upright position on a flat surface. The 40 L column should be lifted from its crate by attaching straps to the lifting eye bolts on the housing.

NOTICE

Do not store the product below 0 °C (32 °F).

6. Getting Started

Set the pressure relief valve to the maximum pressure allowed on the CIM column as indicated in Characteristics of the monolith. Before using the column, an integrity test must be performed. Guideline 'Column integrity test' (biaseparations.com/en/library/guidelines) should be followed. It is advised to repeat this procedure regularly or when deviations in performance are observed.

NOTICE

The column should be equilibrated to working temperature for optimal results. Allow at least 12 h for the column to reach working temperature.

NOTICE

This cGMP compliant column should be sanitised before first use. Follow the sanitisation protocol in this document (see chapter 8.2.) once the 'Column integrity test' has been completed.

6.1. General Recommendations

The following are general guidelines to consider when working with chromatography. The guidelines may not apply to specific column chemistry or sample properties.

- Treat loading material appropriately (e.g. pre-treat, filter, concentrate / dilute, etc.). For more details, please refer to the Guideline 'Pre-treatment of complex biological samples before column purification and regeneration procedures for columns with increased back pressure' (biaseparations.com/en/library/guidelines).
- Always use freshly prepared mobile phases, filtered through 0.2 µm filter, compatible with mobile phases.

- Air bubbles will not disturb the stationary phase and can be washed out of the column. However, drying the monolith risks damaging the stationary phase.
 - Surfactants can improve recoveries in virus purification. Non-ionic surfactants will not interact with ion exchange chromatography media. Non-UV-absorbing (at working wavelengths) surfactants will improve the baseline signal.
 - Ensure all components of the system used are compatible with the working solutions (e.g. sodium hydroxide, organic solvents, high salt concentrations, etc).
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NOTICE

Always ensure mobile phases are compatible before mixing them or applying consecutively on the column. Examples of in-compatible buffers are: magnesium ion-containing buffers and sodium hydroxide (forms precipitate), acetonitrile and sodium hydroxide (forms ammonia and acetate), ammonium acetate and sodium hydroxide (potential formation of explosive atmosphere), ethanol and sodium hydroxide (forms ethoxides). Wash the column with water or another compatible solution when using two incompatible solutions consecutively.

6.2. Buffer Selection

For optimal operation and column lifetime, 10 % (v/v) of organic modifier is recommended in all mobile phases. Exposure to aqueous mobile phases without organic modifier should be kept to a minimum. Consider compatibility between the aqueous (buffering species) and organic (commonly acetonitrile or methanol) portion of the mobile phase, as well as compatibility with the cleaning solution.

Outgassing can occur when organic solvents are combined with aqueous mobile phase. While bubbles do not affect the monolith stationary phase, outgassing could impact the detectors. If the LC system used does not have inline degassing, the mobile phase can be degassed before use. In addition, pre-mixed solutions can be used instead of mixing neat solutions with the LC. For example, to run a gradient from 10 % acetonitrile to 50 % acetonitrile, pre-mix the solutions and run the gradient from 100% A to 100 % B instead of connecting water to inlet A and acetonitrile to inlet B.

The surface of reverse-phase columns are highly hydrophobic, thus excluding water from their surface. To ensure the surface of the stationary phase is suitably "wetted", at least 1 % organic solution should be in the mobile phases used (10 % organic solution recommended in mobile phases).

NOTICE

Organic solvents cause swelling of the stationary phase. Step methods should be avoided, and the concentration of organic solvent should be increased gradually over several column volumes.

NOTICE

Limit contact with aprotic organic solvents (e.g. acetonitrile, DCM), to the method run (gradient). Short (overnight) and long term storage in the designated storage solution. See table Characteristics of the monolith.

NOTICE

Never add acetonitrile directly to alkaline solutions (e.g. NaOH). Acetonitrile hydrolyses to ammonia and acetate.

7. Operating the Column

7.1. Connecting the Column

Position the column with the inlet at the bottom and outlet at the top by placing the 400 | 800 mL column on its stand. The 4000 | 8000 mL columns should stand upright on its wheels. Connect the column to the system with flow turned off in the following order:

1. Carefully remove the blind fitting on the inlet side and connect the inlet tubing.
2. Carefully remove the blind fitting on the outlet side and connect the outlet tubing.

Disconnect by reversing the steps above.

NOTICE

Do not open both inlet and outlet simultaneously to avoid leaking of mobile phase. Changing the order of the above procedure might cause leakage of the mobile phase from the column and affect its performance!

NOTICE

Reversing the flow direction will damage the column. Make sure the column is connected according to the flow direction indicated by the arrow. The 40 L housing has an integrated non-return valve at the column outlet to prevent reversing the flow direction. Do not remove or disassemble the valve. **Note:** Software specific settings which regulate the flow direction should be checked. Ensure the correct flow mode is selected so that flow can go only in the direction indicated on the monolith.

NOTICE

Spikes in pressure generated during sudden pump fluctuations (e.g. immediate application of maximum flow rate or sudden pump stop at high operating pressure) can generate a backpressure shock, which can damage the monolith.

7.2. Equilibration

The column should be equilibrated before use. The equilibration procedure should avoid step transitions between organic and aqueous mobile phases. The binding mobile phase should be of similar composition to the loading sample, and it is recommended to use a binding mobile phase which contains at least low percent of organic solvent (e.g. 1 % acetonitrile).

1. Starting with a column in water or storage solution, wash the column with at least 10 CV of binding mobile phase. **Note:** It is useful to flow the first few CV directly into waste without going through the detector cell. This will remove any air bubbles that may affect the detector cells.
2. Run a linear gradient to 100 % elution mobile phase over 10 CV.
3. Hold at 100% elution mobile phase for 20 CV. A reduced flow rate will ensure extended contact time with elution mobile phase (containing organic solvent) and improve equilibration. Alternatively, a static hold in elution buffer can be used (without flow).

4. Run a linear gradient over 10 CV back to the starting mobile phase.

8. Cleaning | Maintenance

Cleaning and maintenance of the column may improve its lifetime and increase reproducibility. Sample properties should be taken into account for column cleaning.

8.1. Cleaning in Place (CIP)

Sample molecules may bind to the column strongly and not completely elute from the column or may even precipitate on the column. This build-up of contaminants on the monolithic column may cause loss of resolution and binding capacity, increased back pressure, or a complete blockage of the column. A specific CIP procedure should be considered for the type of contaminants present in the sample. An example of a general CIP procedure is presented below.

CAUTION

In case of pressure increase during cleaning, adjust flow rate to remain below the maximum pressure allowed over the column.

Perform the following procedure at up to half the maximum operating flow rate. This will ensure sufficient contact time between the monolith and cleaning solution.

1. Wash the column with 5 CV of deionised water.
2. Wash the column with 5 CV of 1 M NaOH.
3. Wash the column with 5 CV of deionised water.
4. Equilibrate the column.

Note: Beyond NaOH, cleaning with 2-propanol, 0.1 % trifluoroacetic acid (TFA) may be considered. After treatment with NaOH, wash with 5 CV water and then transition to the propanol-TFA buffer in a 10 CV linear gradient.

Note: If CIP does not restore column performance completely, consider performing sanitisation of the column (see below for further instructions).

8.2. Sanitisation

To sanitise the column follow the Cleaning in Place (CIP) Procedure above, extending the contact time with the cleaning solution to 2h. If needed, the column can be disconnected from the system and closed with blind fittings. Re-connect the column and flush with another 5 CV of cleaning solution before proceeding to the next step.

CAUTION

Ensure that the chromatography system and auxiliary components are compatible with NaOH at the concentrations used. Follow all safety instructions when handling NaOH solutions.

9. Storage

Clean and equilibrate the column before storage. It is recommended to store the column in the designated storage solution both overnight and for long-term storage.

NOTICE

Limit contact with aprotic organic solvents (e.g. acetonitrile, DCM), to the method run (gradient). Short (overnight) and long term storage in the designated storage solution. See table Technical Data.

NOTICE

NaOH-ethanol mixtures at any concentration form ethoxide anions that are highly destructive to biomolecules, and ligands on chromatography media. Neutralise the column environment before introducing ethanol.

1. Wash the column with 15 CV storage solution. **Note:** Apply organic solvents using linear gradient of 10 CV or in shallow steps of 10 %. Avoid rapid change from aqueous to organic mobile phase.
2. Seal the column with blind fittings and store at the temperature specified in Technical Data. If there is a possibility of biological contamination from the sample it is recommended to store the column between 2 °C (36 °F) and 8 °C (46 °F).

Note: Reduce the flow rate when using viscous solvents (such as ethanol) to avoid pressure increase.

Note: Never combine acetonitrile and alkaline solutions (e.g. NaOH). Acetonitrile hydrolyses to ammonia and acetate.

10. Troubleshooting

Problems arising during the analysis are usually related to the column, sample, mobile phase, or the instrumentation. It is advisable to use an elimination approach to exclude possible causes. Please refer to our troubleshooting guide (biaseparations.com/en/library/guidelines).

11. Decommissioning | Transportation

If there is reason to return the product, complete a Return Form (biaseparations.com/en/terms-conditions) and contact help@biaseparations.com.

Contaminated samples used during the process that could cause biological or chemical hazards are potentially hazardous substances. If the product has come into contact with hazardous substances, steps must be taken to ensure proper decontamination and declaration.

Procedure

Decontaminate the product. The operator of the product is responsible for adhering to local government regulations on the proper decontamination and declaration for transport and disposal.

12. Ordering Information

Transferring the workflow to a different scale or format (analytical, screening) is simple with CIM™. Contact your local support to find the appropriate products.

Purification Scale Products cGMP Compliant

Catalog number	Product name
901.9001-2	CIMmultus™ SDVB 8 mL cGMP Compliant Monolithic Column (2 µm channels)
914.9001-2	CIMmultus™ SDVB 40 mL cGMP Compliant Monolithic Column (2 µm channels)
911.9001-2	CIMmultus™ SDVB 80 mL cGMP Compliant Monolithic Column (2 µm channels)
924.9001-2	CIMmultus™ SDVB 400 mL cGMP Compliant Monolithic Column (2 µm channels)
921.9001-2	CIMmultus™ SDVB 800 mL cGMP Compliant Monolithic Column (2 µm channels)
934.9001-2	CIMmultus™ SDVB 4000 mL cGMP Compliant Monolithic Column (2 µm channels)
931.9001-2	CIMmultus™ SDVB 8000 mL cGMP Compliant Monolithic Column (2 µm channels)

Purification Scale Products non-cGMP Compliant

Catalog number	Product name
311.9001-2	CIMmultus™ SDVB 1 mL Monolithic Column (2 µm channels)
414.9001-2	CIMmultus™ SDVB 4 mL Monolithic Column (2 µm channels)
411.9001-2	CIMmultus™ SDVB 8 mL Monolithic Column (2 µm channels)
614.9001-2	CIMmultus™ SDVB 40 mL Monolithic Column (2 µm channels)
611.9001-2	CIMmultus™ SDVB 80 mL Monolithic Column (2 µm channels)
814.9001-2	CIMmultus™ SDVB 400 mL Monolithic Column (2 µm channels)
811.9001-2	CIMmultus™ SDVB 800 mL Monolithic Column (2 µm channels)
1014.9001-2	CIMmultus™ SDVB 4000 mL Monolithic Column (2 µm channels)
1011.9001-2	CIMmultus™ SDVB 8000 mL Monolithic Column (2 µm channels)

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The information and figures contained in these instructions correspond to the version date specified below.

Sartorius reserves the right to make changes to the technology, features, specifications and design of the equipment without notice.

Masculine or feminine forms are used to facilitate legibility in these instructions and always simultaneously denote the other gender as well.

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Last updated

06 | 2024

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PSIM-934.9001-2-GMP-2406-ta0